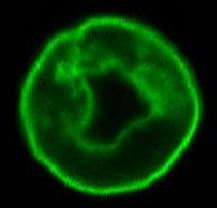




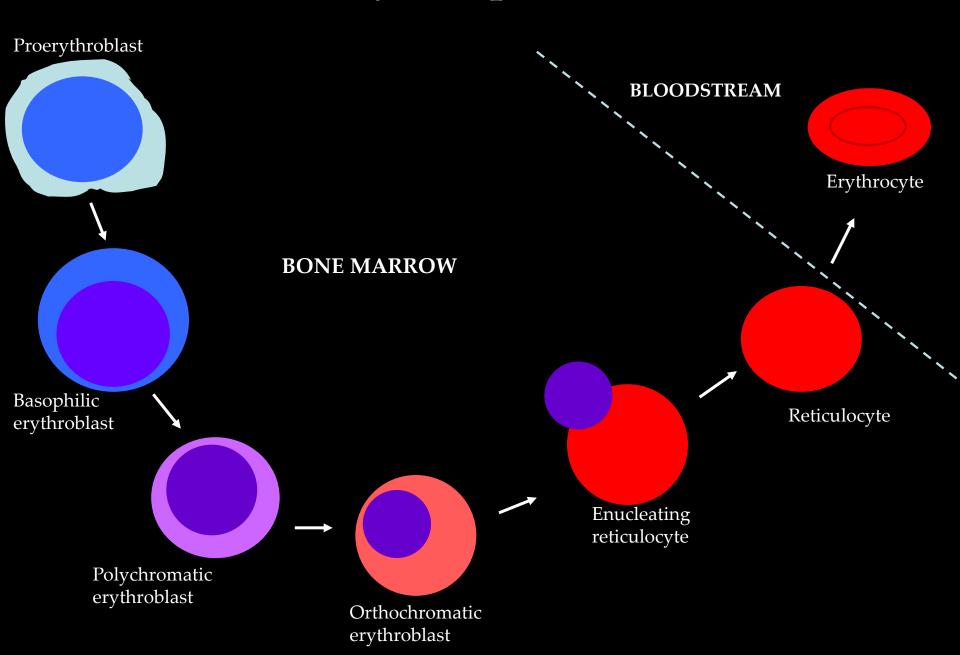
From enucleation to mature erythrocyte: Imaging reticulocyte maturation in vitro



Rebecca Griffiths



Erythropoiesis



Erythroid cultures

Adult peripheral blood CD34+ cells are isolated from waste blood products and cultured in a 3 stage medium.

Overall expansion of cell numbers is $\geq 10^4$ fold.

Enucleation rates are up to 95%.

Leukocyte filtration at the end stage of the culture removes free nuclei, early R1 reticulocytes and normoblasts.

Day 22

R2 Reticulocyte

Day 8

Basophilic

normoblast

Enucleating reticulocyte



Reticulocyte

Day 10

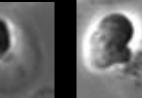
Polychromatic normoblast

Day 6

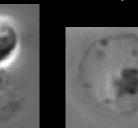


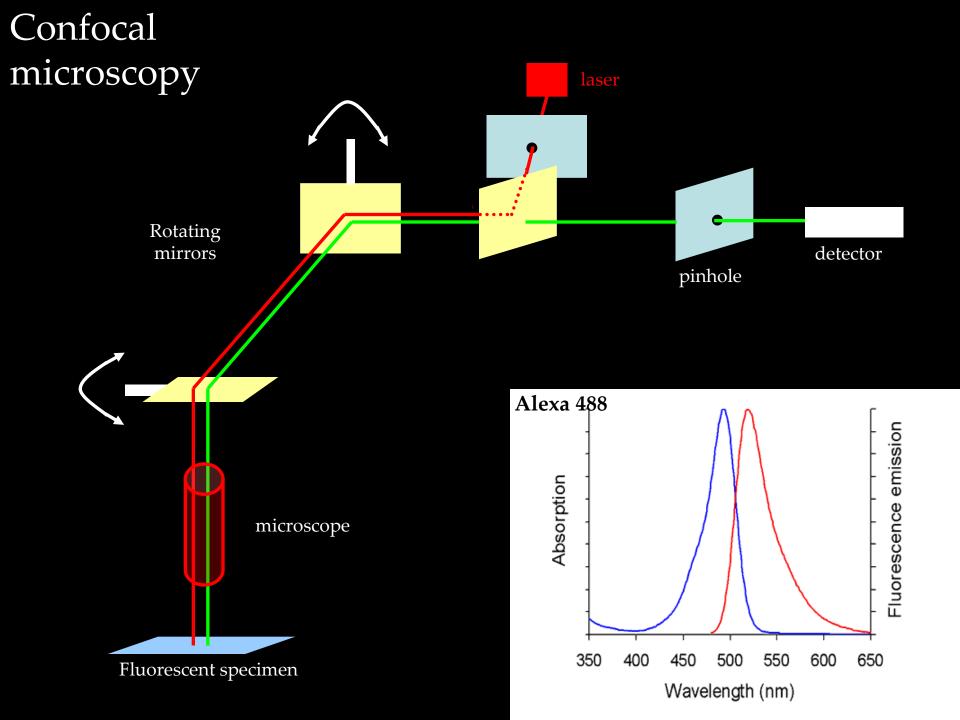
Preproerythroblast

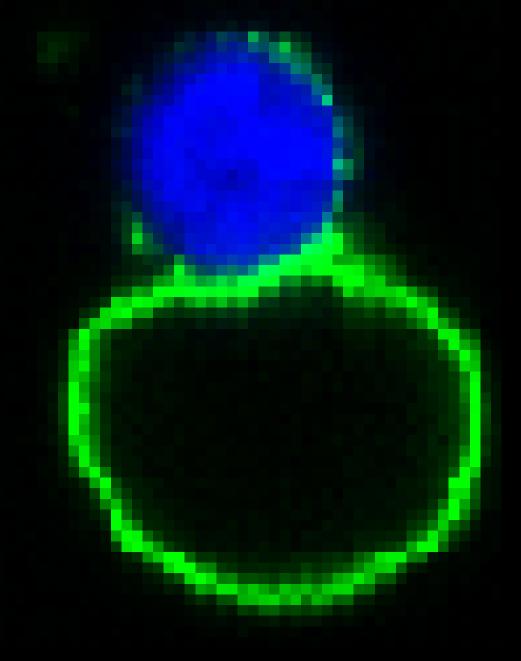
Day 13



Orthochromatic normoblast







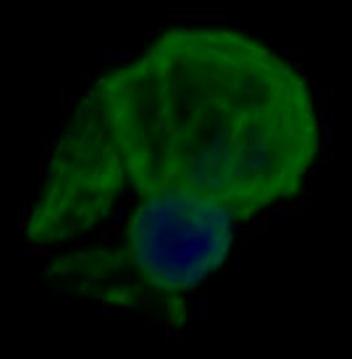
Enucleation

Extrusion of the nucleus at terminal erythroid differentiation – a form of assymetric cytokinesis.

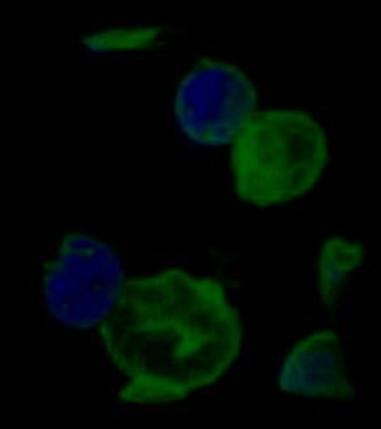
Dynamic state of reorganisation of the plasma membrane and cytoskeleton.

Molecular mechanisms of enucleation are unknown.

GPA & DAPI

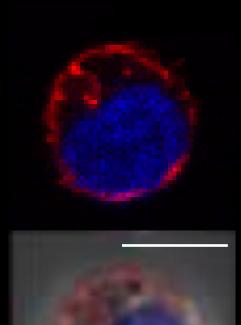


GPA & DAPI



Actin reorganisation at enucleation

erythroblast



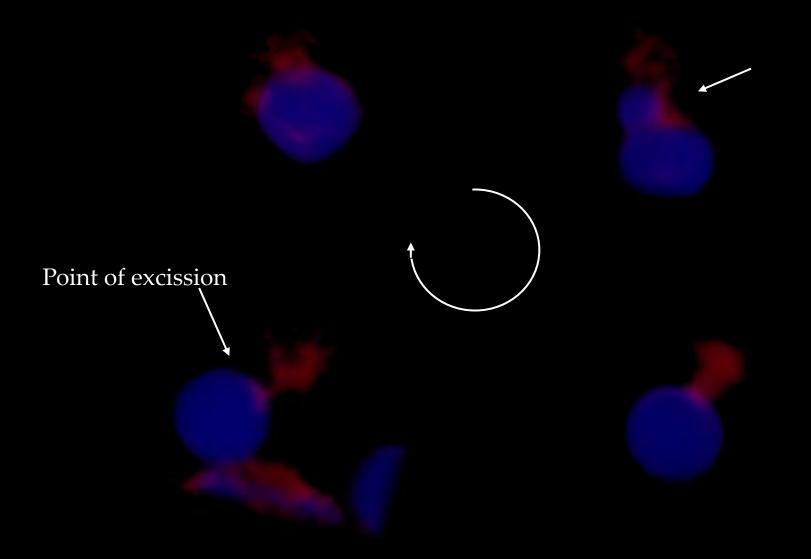
enucleating erythroblast





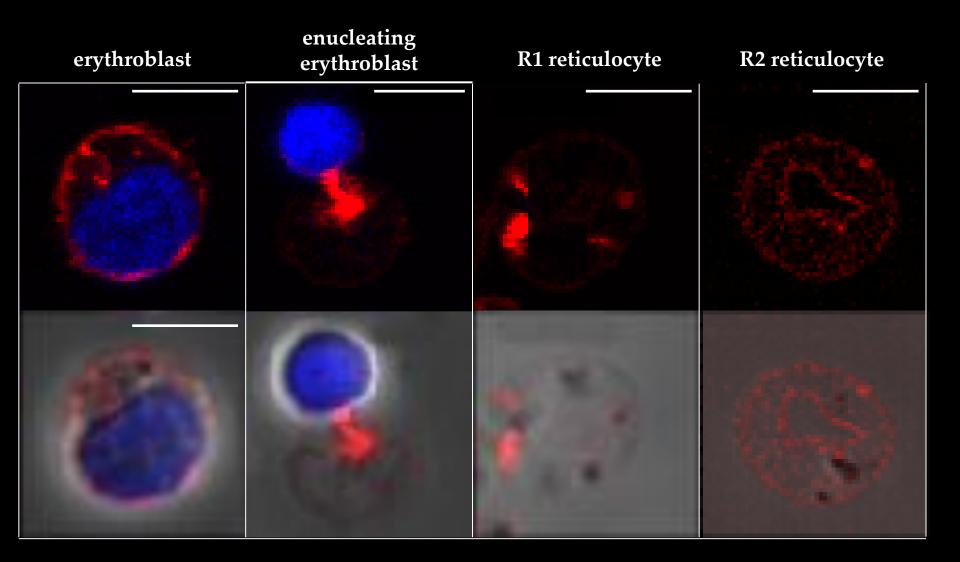
scale bars $5\mu m$

Actin reorganisation at enucleation – the contractile actin ring (CAR) process

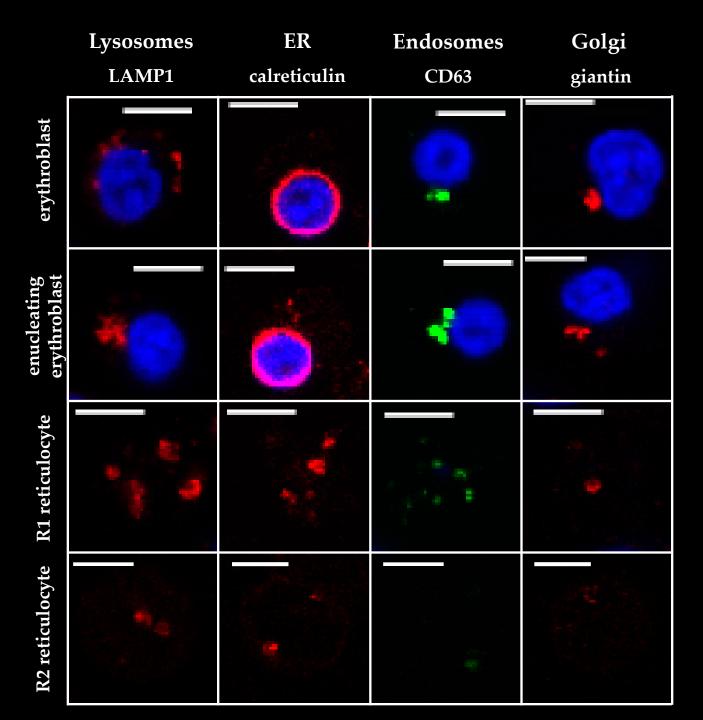


The Contractile Actin Ring

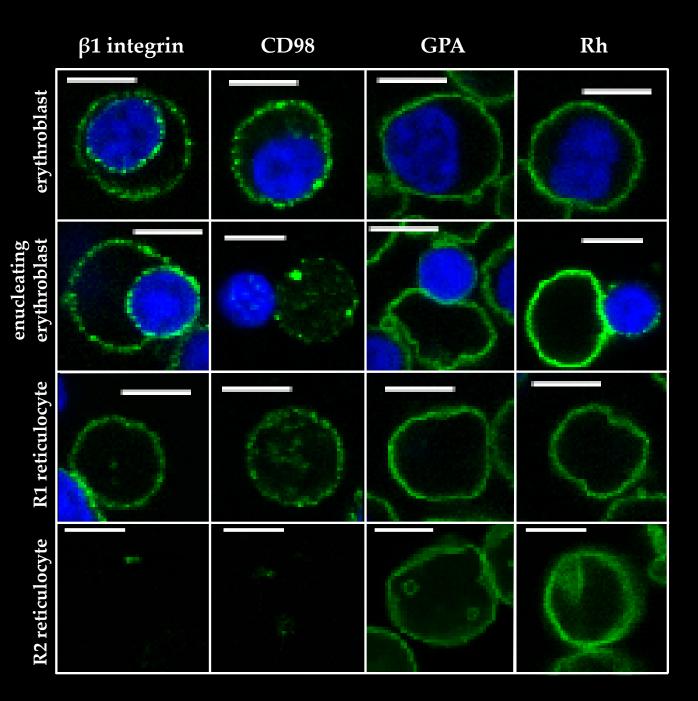
Actin staining in erythropoiesis



Organelle markers at enucleation



Membrane proteins at enucleation

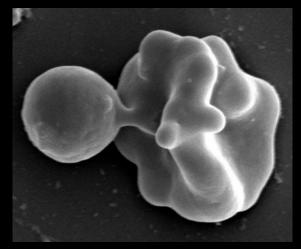


reduction in surface area (20%) and volume

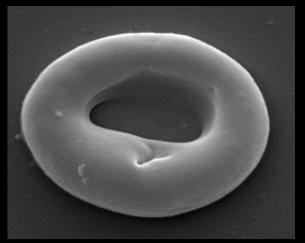
Reticulocyte

degradation / elimination of residual organelles

R1 Motile and multi-lobular



R2 Non-motile



Sabine Kupzig, NHSBT

RBC

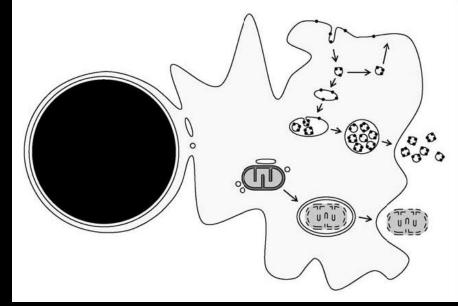
Reticulocyte maturation

Prevailing view

Two distinct mechanisms

Loss of plasma membrane surface area by the endosome-exosome pathway
Degradation / elimination of residual organelles by autophagy

Ney P 2011 Curr Opin Hematol



We have identified a distinct, final stage of reticulocyte maturation . **Griffiths R** *et al* **2012** *Blood*

R2 reticulocytes exhibit large vesicles when stained with GPA

Pre-filtration

GPA and DAP

Post-filtration



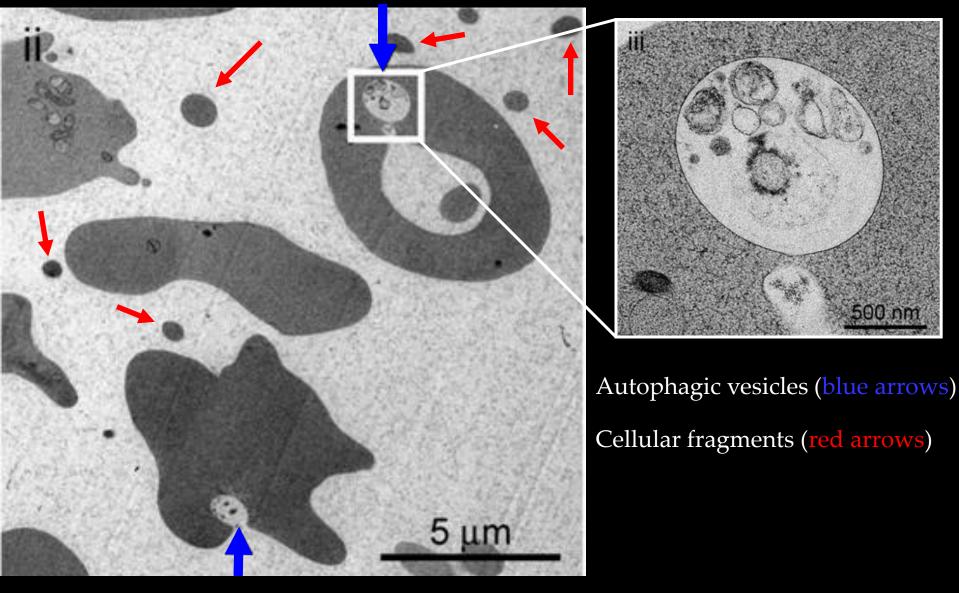
After filtration 64.55% ± 9.37% reticulocytes contain at least one GPA positive vesicle.

> 1.45 µm 8.63 µm

> > 1.45 um

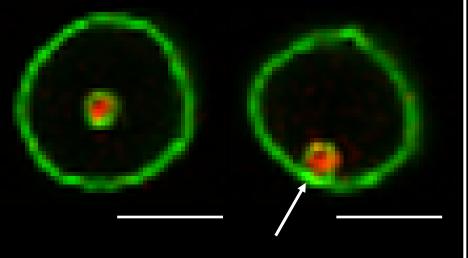
8.63 µm

Transmission Electron Microscopy of Filtered Reticulocytes

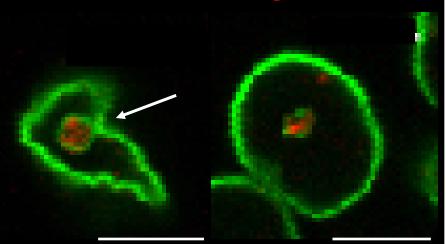


Virginie Betin, University of Bristol

GPA & autophagosome marker LC3

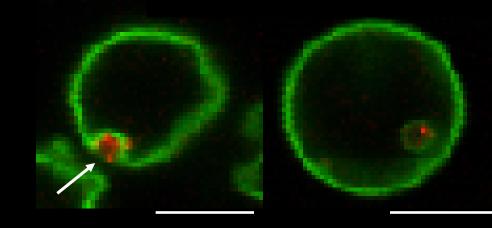


GPA & golgi marker giantin

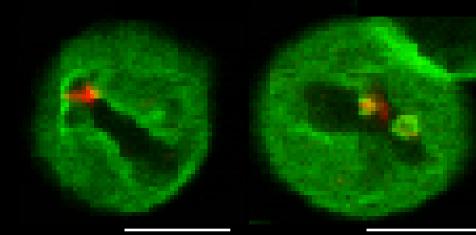


scale bars 5µm

GPA & ER marker calreticulin



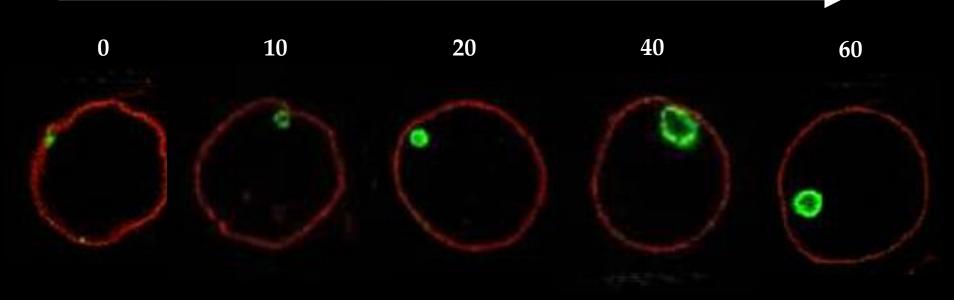
GPA & lysosome marker LAMP1



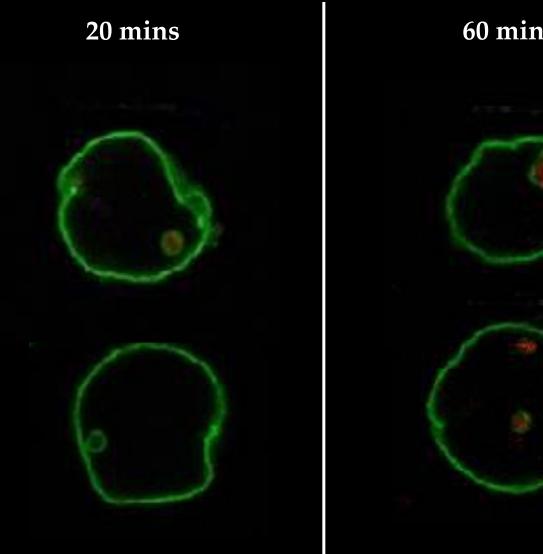
GPA internalisation in Reticulocytes

Internal GPA External GPA

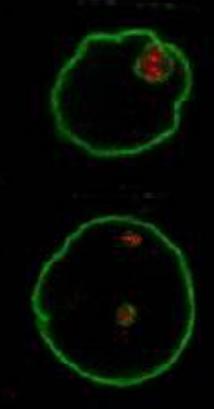
Time (minutes)



Internalised GPA and autophagosome marker LC-3

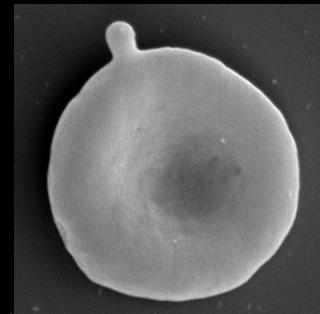


60 mins

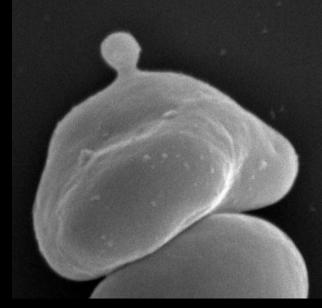


Tosti Mankelow, NHSBT

SEM

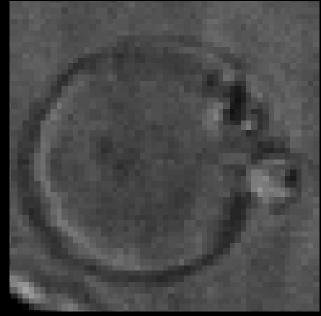


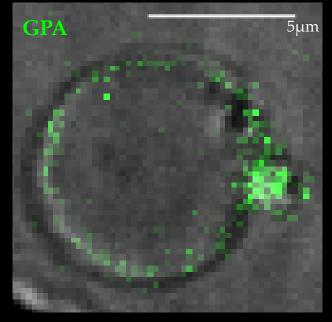
Sabine Kupzig, NHSBT



Blebbing

Confocal (live)

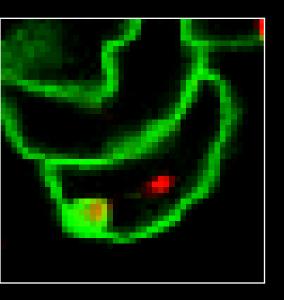




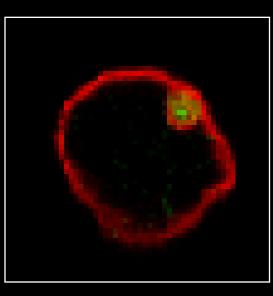
Splenic maturation *in vivo*

Evidence that mature reticulocytes released the bone marrow circulate with autophagic vacuoles and that the final maturation step involving the removal of these vacuoles occurs in the spleen during the first 48-72 hours after release. *In vivo* evidence of GPA positive vesicles - samples from a splenectomised patient

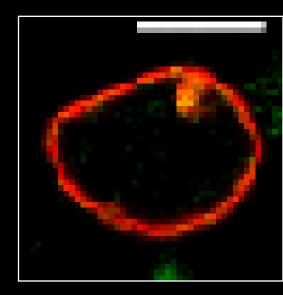
GPA and mitochondrial marker, Mitotracker™



GPA and Golgi marker, giantin



GPA and autophagosome marker, LC3



Conclusions

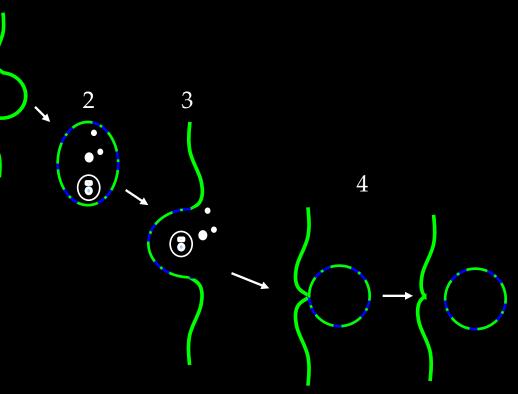
We have identified a distinct, final stage of reticulocyte maturation

1. Large GPA positive vesicles endocytose

2. and fuse with autophagosomes

3. these then fuse with the plasma membrane and the contents exocytose, eliminating unwanted organelles.

4. the membrane then blebs and is released resulting in the loss of plasma membrane surface area.



This is distinct from the exosomal release of plasma membrane proteins (β 1 integrin, CD98 and tetraspanins) seen in earlier R1 reticulocytes.

Our data suggest that reticulocyte maturation occurs in two stages.

Griffiths *et al*. 2012 Blood

Thanks

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University of Bristol

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Jon Lane





