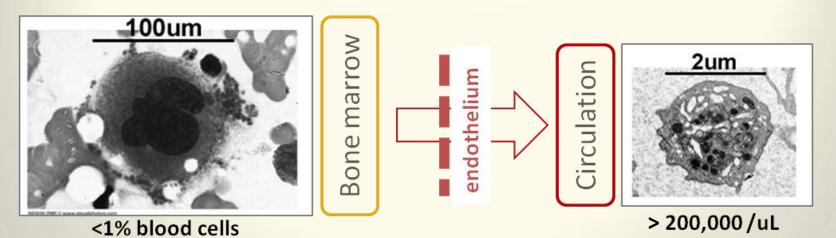
Megakaryocytes from human pluripotent stem cells new perspectives for biological models and clinical applications

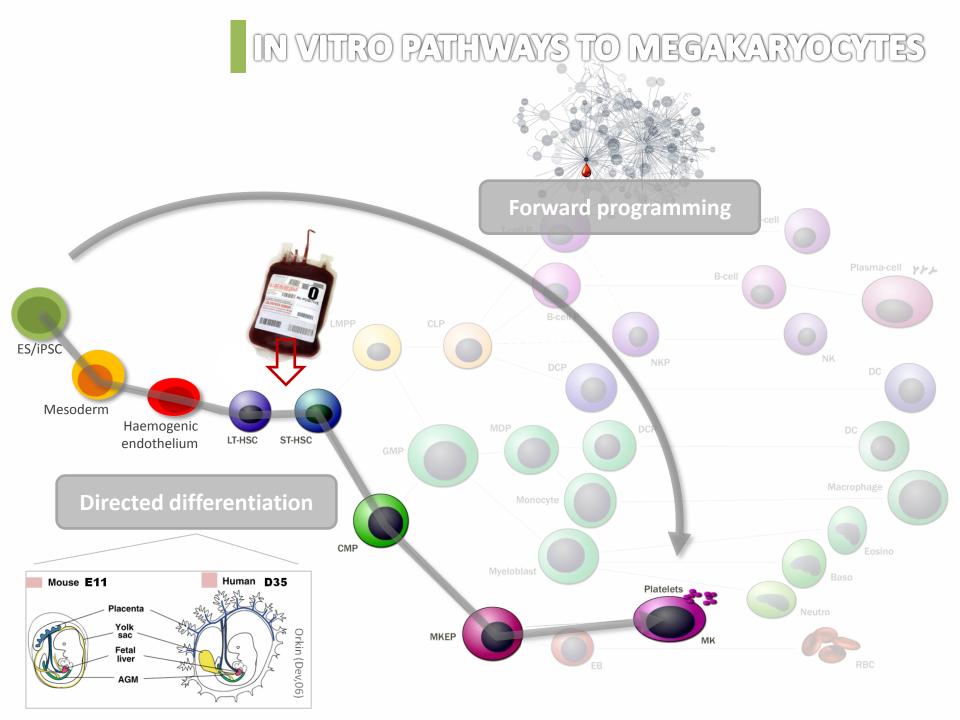
Cedric Ghevaert – Senior Lecturer Transfusion Medicine/ Consultant haematologist Department of Haematology, Division of Transfusion Medicine NHSBT Cambridge Blood Centre



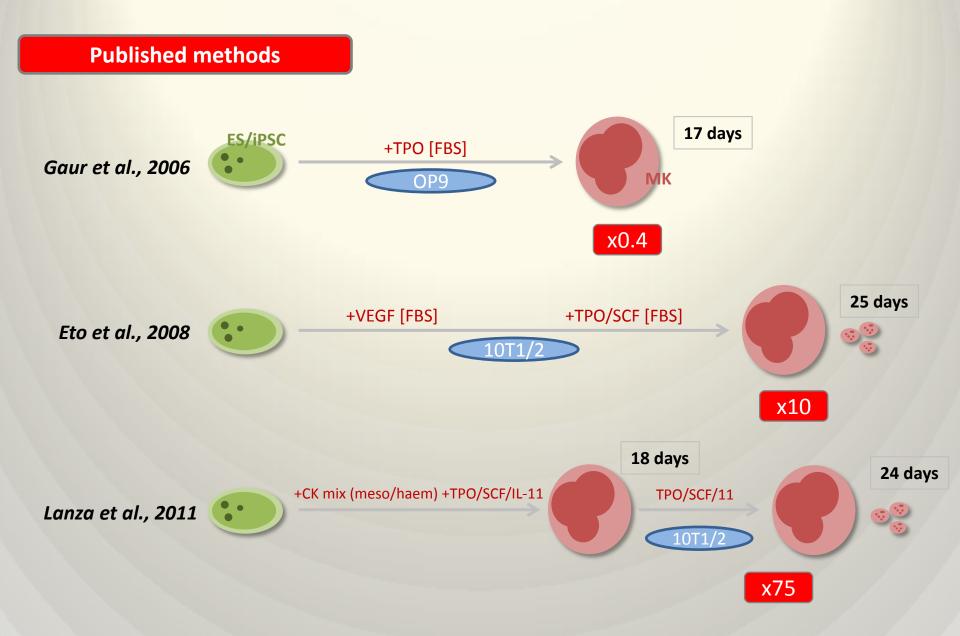
MEGAKARYOCYTE PROFILE



Shaft Branch point Tip Swellings



FROM hipsc to MEGAKARYOCYTES (and platelets?)

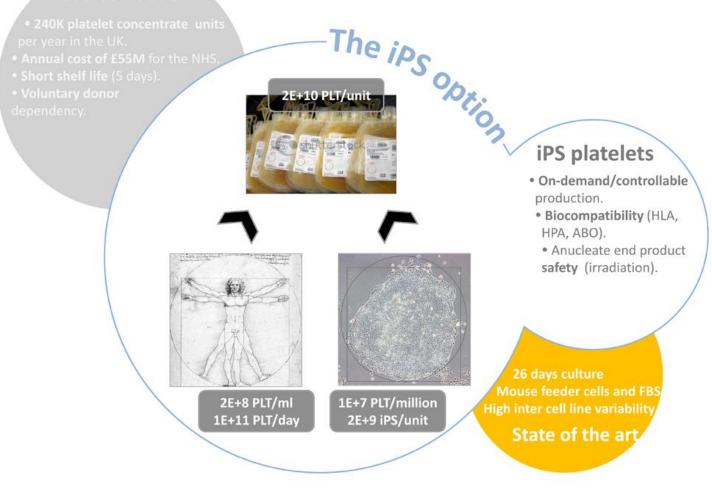


The Long-Term Goal and Drive for This Research

Therapeutic application

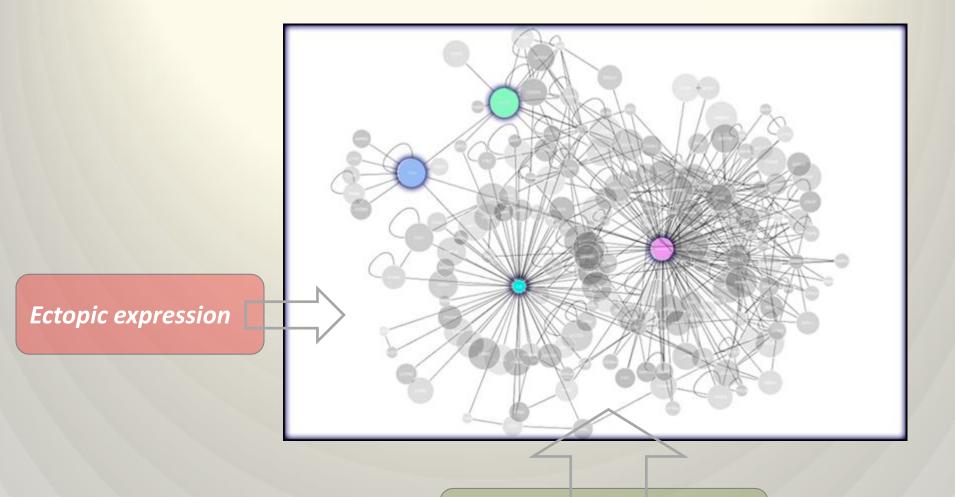
In vitro production of platelet for transfusion

Platelet needs



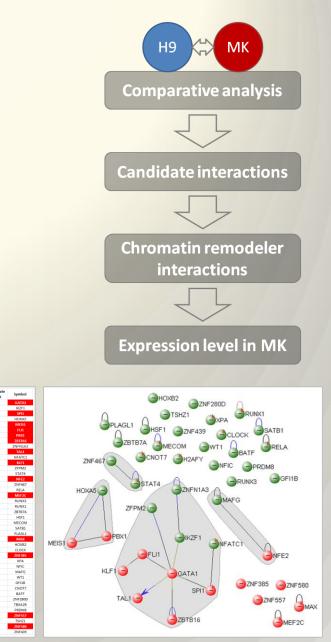
MK PROGRAMMING APPROACH HYPOTHESIS

Ectopic expression of master transcription factors to impose a MK phenotype to unrelated cell types

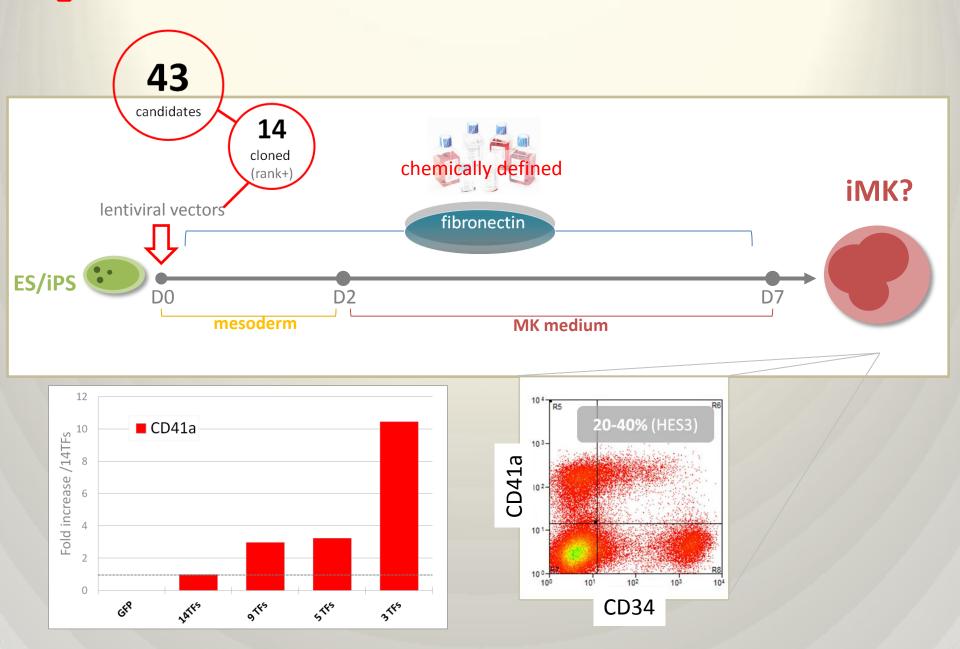


Environmental stimuli

MK Forward Programming – 1st STEPS



MK Forward Programming – 1st STEPS



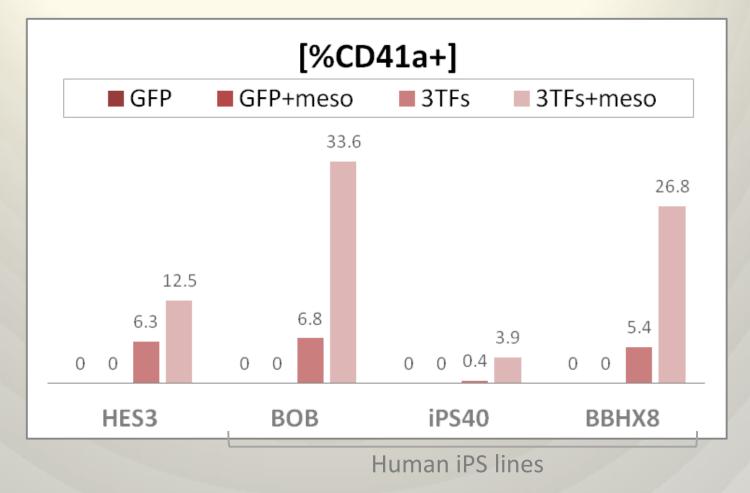
MK Forward Programming – 1st STEPS

0.15 FLI1 1.56 MPL NFE2 0.21 gene expression /HMBS gene expression /HMBS xpression /HMBS Gene expression in sorted populations 0.08 0.02 0.00 ■ GATA1 ■ MPL ■ NFE2 ■ ZFPM1 ■ MEIS1 ES GFP **3TFS** ES expression /MDH1 ZFF GATA1 0.16 2.86 gene expression /HMBS gene expression /HMBS gene 0.01 0.03 0.00 0.49 ES GFP **3TFS** ES 0.14 0.15 0.06 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.000.16 TAL1 RU CD41-CD34-CD41-CD34+ CD41+ gene expression /HMBS gene expression /HMBS gene expression /H 0.08 0.06 0.07 0.03 0.00 0.00 **3TFS** ES GFP **3TFS** GFP **3TFS** ES GFP ES early late

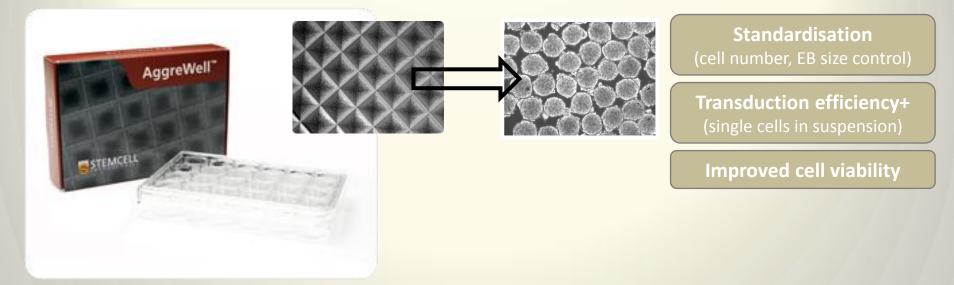
whole population D7

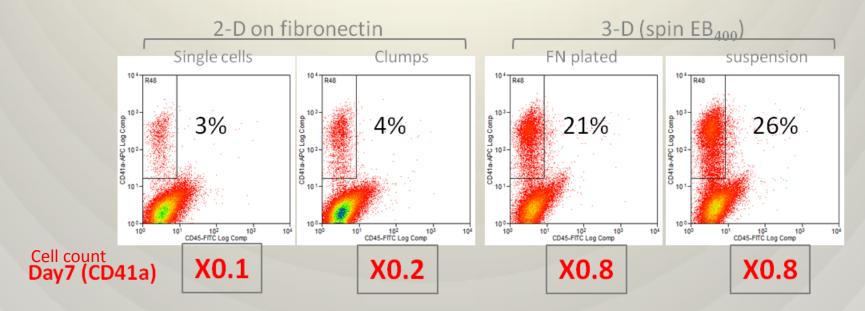
DEVELOPMENT - MESODERM INDUCTION





DEVELOPMENT – GOING 3D

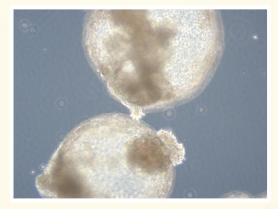


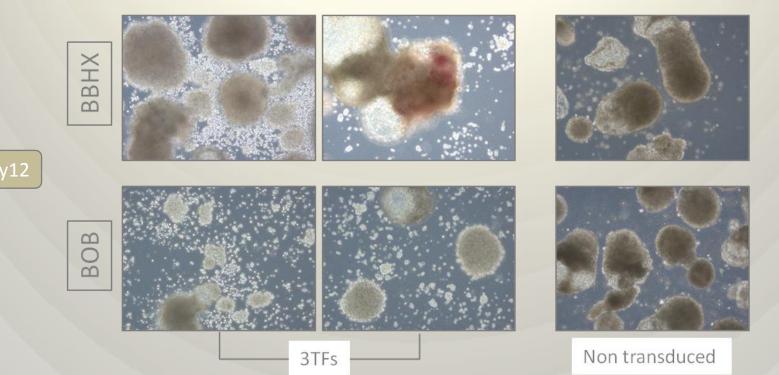


DEVELOPMENT – GOING 3D

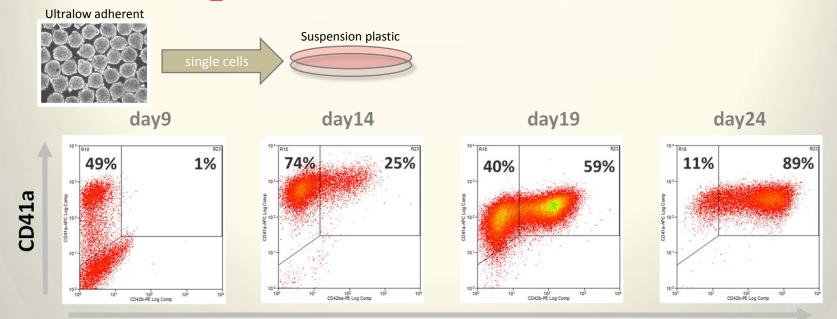




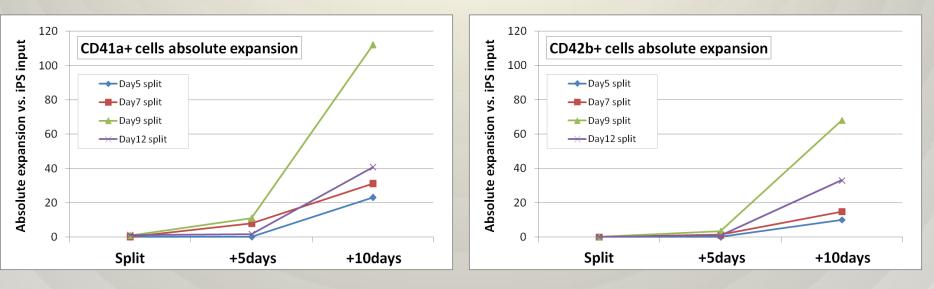




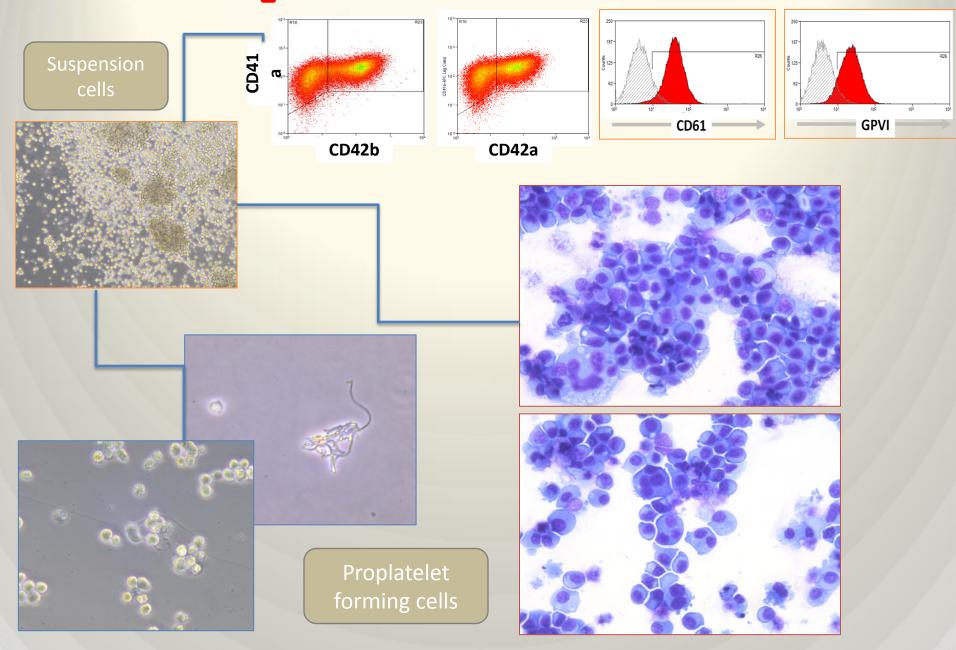
FOP MK MATURATION – 2nd CULTURE STEP



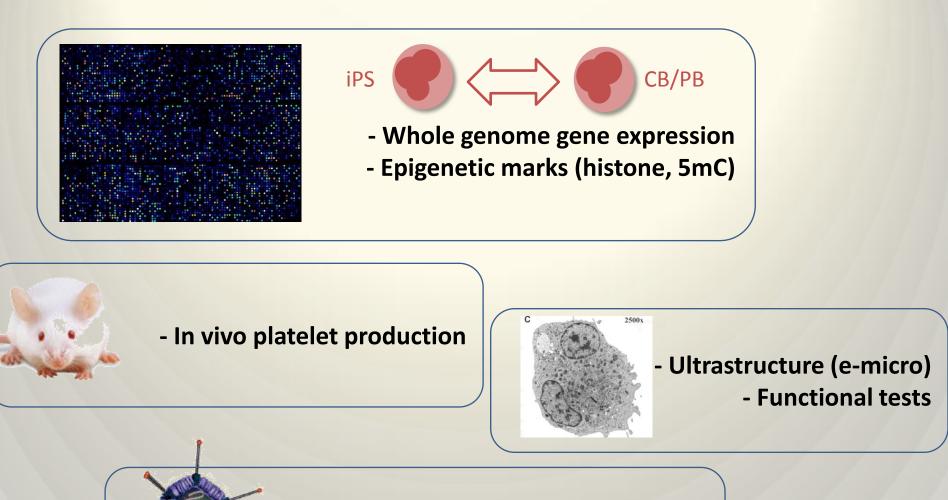
CD42b



FOP MK MATURATION – 2nd CULTURE STEP



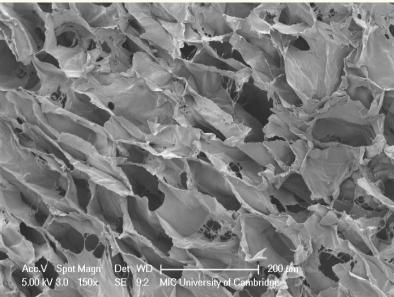
MK FORWARD PROGRAMMING - ONGOING...

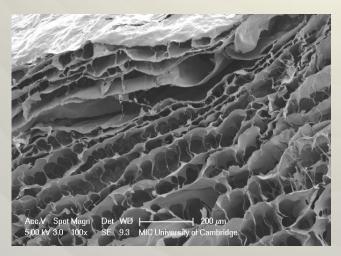


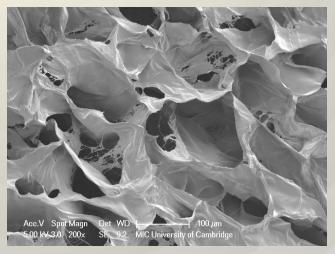
- Non integrative forward programming

Scaling up and promoting platelet production

COLLAGEN SCAFFOLDS

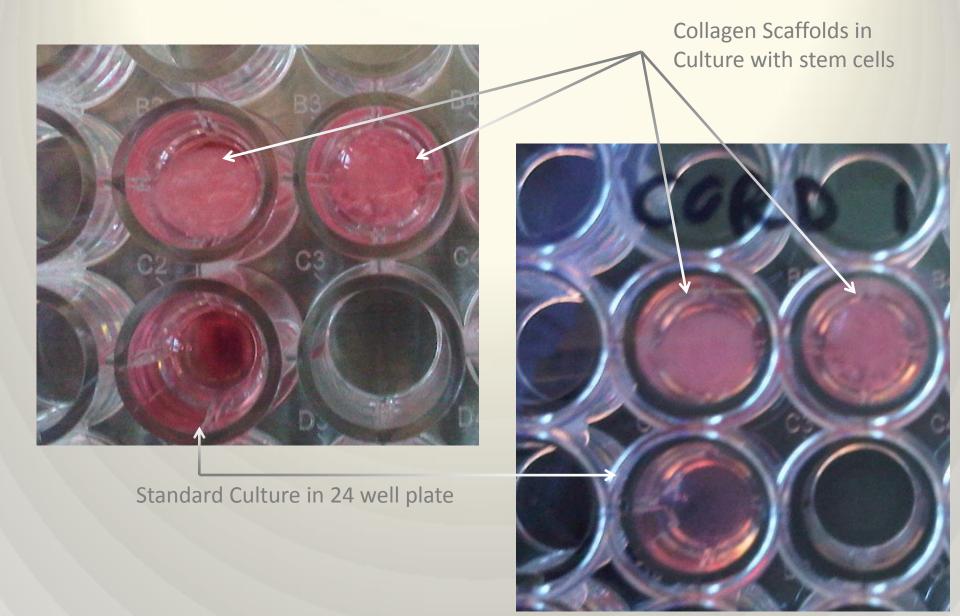




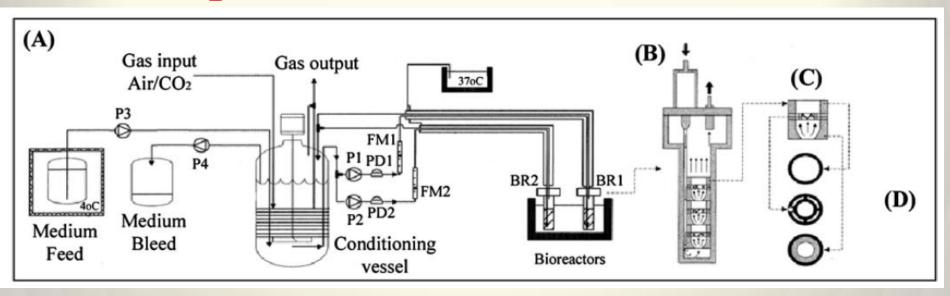


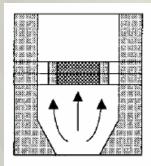
Variation of freeze dry conditions to provide different pore size and architecture

Scaling up and promoting platelet production



Scaling up and promoting platelet production

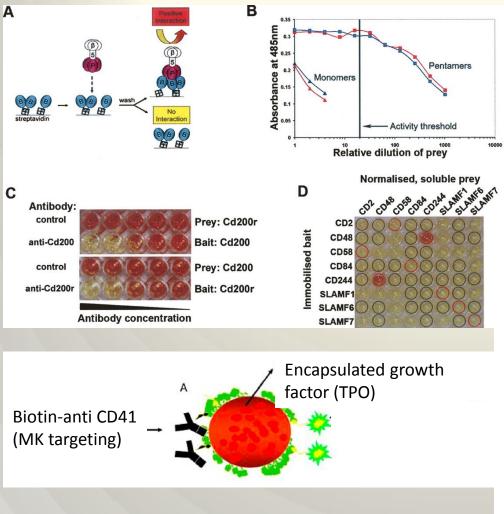






MK Forward Programming - FUNCTIONALISING SCAFFOLDS **DELIVERING CYTOKINES**

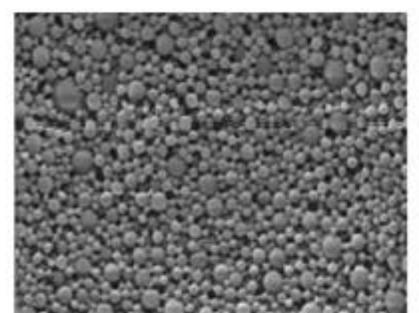




Steenblock et al., Molecular Therapy 2008

Bushell et al., Genome Res 2009

TARGETED NANOBEADS



CONCLUSIONS...

MK FORWARD PROGRAMMING OUTCOME

- **Cells expressing key features of mature megakaryocytes** have been obtained in chemically defined conditions in 19 days from several human ESC and iPSC lines
- The numbers of cells obtained is superior to any other protocol and compatible with the goal of in vitro production of platelets for human use.

SHORT TERM PERSPECTIVES

- Transcriptome analysis (+ epigenetic profiling) vs. cord blood derived MKs
- Functional assays (*in vitro* proplatelet formation, *in vivo* transplant studies in NSG mice)
- Test of additional candidate TFs to improve MK maturation step

MEDIUM TERM PERSPECTIVES

Biological models

Genetic engineering of iPSC lines for MK/PLT biology studies

Tagging of novel MK genes for gene function studies

MK targeted expression of maturation factors

Disease modelling

Derivation of iPSC lines from patients with inherited platelet disorders

TAR and GPS follow up studies

Systems biology

Large scale genetic and epigenetic characterization of iPSC lines and derived MK

Explore haematopoietic/MK potential inter line variability

identify genetic and epigenetic factors controlling megakaryopoiesis from iPSC

First quality assessment of a realistic iPSC derived cell therapy product, i.e. MK/PLT

ACKNOWLEDGMENTS

